

Role of *Saccharomyces* Probiotic in Addressing Heat Stress in Broiler Chickens

* Abdelalim F Abd elalim * Naglaa Z.H.Eleiwa ** Arafa M.M ** Abd Elfattah I. El-Zanaty **Mohamed.A.Abo Elfadl

*Pharmacology Department, Faculty of Veterinary Medicine, Zagazig University, 44519, Egypt;

** Animal Health Research Institute, Dokki, Egypt, (AHRI-ARC).

Application of natural feed additives with a view to enhance production performance, carcass quality and state of health has constituted an important request in production of poultry especially under heat stress conditions. This study aimed to investigate the impacts of dietary supplementation of *Saccharomyces cerevisiae* on growth performance parameters, some blood biochemical findings, antioxidant status in growing broilers exposed to heat stress. For this purpose, a total of 80 one-day-old chicks were randomly allotted into 4 dietary groups with 20 chicks each, which were fed for 35 days with a basal diet., Group1 served as the negative control group unexposed to heat stress (21-22 °C). While, Group 2 served as the positive control group exposed to heat stress (33-35 °C).While, Groups 3 un exposed to heat stress and supplemented with *Saccharomyces cerevisiae* (SC) and Groups 4 served as experimental groups exposed to heat stress (33-35 °C) and supplemented with *Saccharomyces cerevisiae* (SC). respectively The obtained results showed that SC dietary supplementation to basal diet in groups T3, T4 resulted in improved growth performance parameters (increased final B.W, BWG, F.C and F.C.R), improved liver and kidney function (reduced serum AST, ALT, creatinine and uric acid level), improved protein level (total protein& albumin & globulin), improved antioxidant status (reduced serum MDA levels, Cortisol and increased serum CAT, SOD, TAC and GPx activities), increase antibody titer against IB& ND, decrease level of glucose, PH, Haptoglobin, CRP. as compared to birds in group 2.

Keywords: Heat stress, Growth performance, Antioxidant capacity, *Saccharomyces cerevisiae*.

Introduction:

The poultry industry is growing across the world to fulfill the increasing demands of poultry meat and eggs. Poultry birds play a significant role in boosting the economy of many developing countries, providing nutrition and also contributing to the livelihood of many, particularly in rural areas (**Demeke 2004**).

The poultry have a great contribution to worldwide nutrition and food safety. It aids in the production of reasonably priced protein, important micronutrients, and energy to human beings (**Mottet and Tempio, 2017**).

In recent times, the environmental temperature increase globally becomes apparent severe challenge attacking the chicken farming in tropical and subtropical areas. In poultry, the heat injury happened if the ambient environmental temperature surpasses the thermo-neutral zone (**El-Kholy et al., 2018**).

Heat stress (HS), one of the most serious climate problems of tropical and subtropical regions of world, negatively affects the production performance of poultry and livestock. Concisely, HS is characterized by endocrine disorders, reduced metabolic rate, lipid peroxidation, decreased feed consumption, decreased BW gain, higher feed conversion ratio (FCR), immunosuppression, and intestinal microbial dysbiosis (**Sohail et al., 2011**).

Heat stress also increases peroxidation of lipid through increasing the free radical's generation. Peroxidation of lipid is also a great reason for deterioration of quality of meat, affecting texture, flavour, colour and nutritional benefits (**Botsoglou et al., 2010**).

Heat stress results from a negative balance between the net amounts of energy flowing from the animal's body to its surrounding environment And the amount of heat energy produced by the animal. This imbalance may be caused by variations of a combination of environmental factors

(e.g: sunlight, thermal irradiation, and air temperature, humidity and movement) and characteristics of the animal (e.g: species, metabolism rate and thermoregulatory mechanisms). Environmental stressors, such as heat stress, are particularly detrimental to animal agriculture (**Nienaber et al., 2007–Renaudeau et al., 2012**).

A recent study (**Felver et al., 2014**) showed that birds subjected to heat stress conditions spend less time feeding, more time drinking and panting, as well as more time with their wings elevated, less time moving or walking, and more time resting.

Animals utilize multiple ways for maintaining thermoregulation and homeostasis when subjected to high environmental temperatures, including increasing radiant, convective and evaporative heat loss by vasodilatation and perspiration (**Yousaf et al., 2019**).

Probiotics are live microbes that are given as food additives and provide benefits to the host by improving the balance of intestinal microbial populations (**Abudabos et al. 2016**).

Two basic Mechanisms by which probiotic act to maintain a beneficial microbial population include competitive exclusion and immune modulation. Competitive exclusion involves competition for substrates, production of antimicrobial metabolites that inhibit pathogen and competition for attachment sites (**Yang et al., 2009**), by directly interacting with gut mucosal immune system, probiotics can modulate either innate or acquired immunity (**Dugas et al., 1999**). Further, specific immune modulatory effects of probiotics are dependent on the strain or species of bacteria included in the probiotics (**Edens, 2003 and Huang et al., 2004**).

In the last decade, the *Saccharomyces cerevisiae* (SC) yeast has received great attention. Supplementation of feeds with living yeast cells has been reported to improve digestibility of feed, enhance feed efficiency and performance of animal, decrease the pathogenic bacteria number, enhance the health of animal and decrease the pad environmental effects on livestock production (**Elghandour et al., 2019**).

Saccharomyces cerevisiae is arguably one of the most studied microorganisms. However, its mechanism of action as a probiotic has not been fully understood (**Ahmad., 2006**). It has been reported that it elaborates enzymes which aid in digestion and produces lactic acid which increases acidity and reduces pH of the digestive tract, thereby preventing the growth of pathogenic organisms and aiding in enzymatic activity. This increases digestibility and utility of minerals, proteins, and amino acids (**Gunal et al. 2006**).

Saccharomyces cerevisiae has mannan oligosaccharide (MOS) in its cell wall which is a natural feed additive that is involved in helping the growth of beneficial microbes and inhibiting that of the pathogenic organisms (**Yang et al. 2008**).

2- Material & Method:

Dietary treatments

The broilers feeding programme consisted of starter (0–21 day) and finisher (21–35 day) basal diets that were processed to supply dietary nutrient requirements of the birds (NRC, 1994). Each basal diet was classified into 4 Groups, which were prepared as follows: G1: a negative control, basal diet with no supplementation without heat stress (21 -22 °C) exposure; G2: positive control, basal diet with no supplementation with heat stress (30 - 35 °C), G3: basal diet un exposed to heat stress and supplemented with *Saccharomyces cerevisiae* (SC), G4: basal diet under heat stress supplemented with *Saccharomyces cerevisiae* (SC).

Management of birds and experimental design

The present experiment was conducted on 80, healthy one day old unsexed broiler (Arbor Ecars) chicks purchased from El-kahera Poultry Company. The chicks were randomly allocated into 4 groups. The broiler chicks were fed on well balanced ration. Starter diet was given till the 20 day of age after the chick were fed on finisher diet which was given from the 21 day till the end of the experiment. Water & food were offered ad libitum. The chicks were housed in clean well ventilated separate experimental rooms, previously fumigated with formalin & potassium permanganate. The floor was bedded by fresh clean chopped wheat straw forming litter of 3.5cm depth. Each room was provided by suitable numbers of feeders & water utensils. The chicks were vaccinated against most common viral diseases which infect the broiler chicks.

Ethical statement

All chickens included in this study were handled according to the regulations of the Animal Ethics of Institutional

Animal Care Committee (ARC-IACUC) at Agriculture Research Center, Egypt (Approval Number: ARC-AH-22-14), with good animal practice following the guidelines of the research code of ethics (ARC-IACUC) at Agriculture Research Center. All efforts were done to minimize painful sense and discomfort to the birds.

Probiotic:

Saccharomyces was obtained from biotechnology department at Animal Health Research Institute, Dokki, Giza, Egypt (ARC-AHRI).

Dose: 2.5×10^7 CFU/g ration (Aalaei et al., 2018).

Route: on ration.

Blood sample(serum):

Serum was collected from 5 birds at the 15th & 35th days of ages in plain clean well dried centrifuge tube used for separation of serum to be used in estimation of biochemical parameters & serological studies.

Statistical Analysis:

Data statistically analyzed & each reading represents means $\pm$ standard deviation. The statistical evaluation of all data done using one-way analysis of variance (ANOVA) followed by Dunnett's test using a computer program (Statistical Package for Social science, version 16) P value ≤ 0.05 & P value ≤ 0.001 regarded as statistically significant SPSS program (2008).

Results:

1--Effect of *Saccharomyces* on growth performance

a) Body weight (B.W)

The effect of supplementation of *Saccharomyces*'s on the B.W in broiler chicken is presented in Table (1)

Feeding of chicken on diet supplemented with *Saccharomyces*'s (2.5×10^7 CFU/g) induced a significant ($P < 0.05$) increase in body weight compared with control group all over the experimental period.

Chicks experimentally exposed to heat stressed and fed on basal diet evoked a significant ($P < 0.05$) decrease in body weight all over the experimental period.

Feeding of heat stressed chicken on diet supplemented with *Saccharomyces*'s (2.5×10^7 CFU/g) displayed a significant ($P < 0.05$) increase in Body weight compared with heat stressed group all over the experimental period.

b) B.W.G

The effect of supplementation of *Saccharomyces's* on the B.W.G in broiler chicken is presented in Table (1)

Feeding of chicken on diet supplemented with *Saccharomyces's* (2.5×10^7 CFU/g) induced a significant ($P < 0.05$) increase in body weight gain compared with control group all over the experimental period.

Chicks experimentally exposed to heat stressed and fed on basal diet evoked a significant ($P < 0.05$) decrease in body weight gain all over the experimental period.

Feeding of heat stressed chicken on diet supplemented with *Saccharomyces's* (2.5×10^7 CFU/g) displayed a significant ($P < 0.05$) increase in body weight gain compared with heat stressed group all over the experimental period.

c) Food consumption (F.C):

The effect of supplementation of *Saccharomyces's* on the F.C in broiler chicken is presented in Table (2)

Feeding of chicken on diet supplemented with *Saccharomyces's* (2.5×10^7 CFU/g) induced a significant ($P < 0.05$) increase in food consumption compared with control group on 15th days of age and significant ($P < 0.05$) decrease on 35st days of age.

Chicks experimentally exposed to heat stressed and fed on basal diet evoked a significant ($P < 0.05$) decrease in food consumption all over the experimental period.

Feeding of heat stressed chicken on diet supplemented with *Saccharomyces's* (2.5×10^7 CFU/g) displayed a significant ($P < 0.05$) increase in food consumption compared with heat stressed group all over the experimental period.

D) Feed conversion ratio (F.C.R):

The effect of *Saccharomyces's* on F.C.R of heat stress exposed chicken was illustrated in table (2)

Feeding of chicken on diet supplemented with *Saccharomyces's* (2.5×10^7 CFU/g) induced non-significant ($P < 0.05$) change in food conversion ratio compared with control group on 15th days of age and significant ($P < 0.05$) decrease on 35st days of age.

Chicks experimentally exposed to heat stressed and fed on basal diet evoked a significant ($P < 0.05$) increase in food conversion ratio on 15th days of age and significant ($P < 0.05$) decrease on 35st days of age.

Feeding of heat stressed chicken on diet supplemented with *Saccharomyces's* (2.5×10^7 CFU/g) displayed a significant ($P < 0.05$) decrease in food conversion ratio compared with heat stressed group all over the experimental period.

Table (1): The effect of *Saccharomyces's* on body weight and body weight gain in broiler chicken exposed to heat stress on the 15th and 35st days of age.

Days Groups	15 th			35 st	
	Initial B.W	B.W. (gm)	B.W.G (gm)	B.W. (gm)	B.W.G (gm)
Group 1	70.86±1.79	386.00±4.30 ^{de}	315.4±4.65 ^{ed}	1823.6±66.97 ^e	1437.6±66.43 ^d
Group 2	72.88±1.62	244.4±4.69 ^f	171.52±3.53 ^f	1493.4±34.61 ^f	1249.0±30.88 ^e

Group 3	73.58±1.66	421.8±5.66 ^{ab}	348.22±7.06 ^{ab}	2023.4±8.63 ^c	1601.6±9.78 ^c
Group 4	73.64±1.25	394.4±4.30 ^{cd}	320.36±4.76 ^{cd}	1919.6±24.49 ^d	1525.6±23.68 ^c

Table 2: The effect of *Saccharomyces*'s on food consumption and food conversion ratio in broiler chicken exposed to heat stress on the 15th and 35st days of age.

Days Groups	15 th		35 st	
	FC	FCR	FC	FCR
Group 1	1237.8±19.22 ^c	3.53±0.42 ^c	1816±10.29 ^a	1.272±0.05 ^a
Group 2	1086.8±12.40 ^d	6.332±0.16 ^a	1438±20.34 ^e	1.1536±0.03 ^{ab}
Group 3	1306.4±6.23 ^{ab}	3.75±0.07 ^c	1763±8.60 ^{ab}	1.097±0.01 ^{abc}
Group 4	1304.4±3.41 ^{ab}	4.01±0.06 ^{bc}	1646±22.93 ^d	1.078±0.01 ^{abc}

Letter differ at $p < 0.05$.

2--Effect of *Saccharomyces*'s on liver function test (ALT& AST)

The effect of *Saccharomyces*'s on ALT& AST of heat stress exposed chicken was illustrated in table (3)

Feeding of chicken on diet supplemented with *Saccharomyces*'s (2.5×10^7 CFU/g) induced significant ($P < 0.05$) decrease in ALT & AST compared with control group all over the experimental period.

Chicks experimentally exposed to heat stressed and fed on basal diet evoked a significant ($P < 0.05$) increase in ALT & AST all over the experimental period.

Feeding of heat stressed chicken on diet supplemented with *Saccharomyces*'s (2.5×10^7 CFU/g) displayed a significant ($P < 0.05$) decrease in ALT & AST compared with heat stressed group all over the experimental period.

Table 3: The effect of *Saccharomyces*'s on liver function test in broiler chicken exposed to heat stress on the 15th and 35st days of age.

Days Groups	15 th		35 st	
	ALT (µl/ml)	AST (µl/ml)	ALT (µl/ml)	AST (µl/ml)
Group 1	74.6±0.92 ^c	99.8±2.15 ^c	78.6±1.20 ^b	109.4 ±0.87 ^b
Group 2	93.6±2.01 ^a	231±9.92 ^a	95.2±1.85 ^a	238.6 ±0.92 ^a
Group 3	68.8±0.58 ^d	96.8±0.66 ^d	63±0.94 ^e	89.4 ±1.46 ^c
Group 4	80.2±1.06 ^b	124±2.55 ^b	74.2±1.49 ^c	107.2 ±1.93 ^b

Letter differ at $p < 0.05$.

3--Effect of *Saccharomyces's* on kidney function test (creatinine & uric acid)

The effect of *Saccharomyces's* on creatinine & uric acid of heat stress exposed chicken was illustrated in table (4)

Feeding of chicken on diet supplemented with *Saccharomyces's* (2.5×10^7 CFU/g) induced significant ($P < 0.05$) decrease in creatinine & uric acid compared with control group all over the experimental period.

Chicks experimentally exposed to heat stressed and fed on basal diet evoked a significant ($P < 0.05$) increase in creatinine & uric acid all over the experimental period.

Feeding of heat stressed chicken on diet supplemented with *Saccharomyces's* (2.5×10^7 CFU/g) displayed a significant ($P < 0.05$) decrease in creatinine & uric acid compared with heat stressed group all over the experimental period.

Table 4: The effect of *Saccharomyces's* on Kidney function test in broiler chicken exposed to heat stress on the 15th and 35st days of age.

Days Groups	15 th		35 st	
	Creatinine (µl/ml)	Uric acid (µl/ml)	Creatinine (µl/ml)	Uric acid (µl/ml)
Group 1	0.604±0.03bc	2.86±0.06bcd	0.772±0.02bc	3.52±0.15d
Group 2	1.08±0.04a	5.52±0.48a	1.244±0.03a	7.32±0.27a
Group 3	0.518±0.03cd	2.4±0.10def	0.68±0.02d	3.16±0.05de
Group 4	0.648±0.04b	3.14±0.12bc	0.794±0.01b	4.00±0.12bc

Letter differ at $p < 0.05$.

4--Effect of *Saccharomyces's* on protein level (total protein & albumin & globulin)

The effect of *Saccharomyces's* on total protein & albumin & globulin of heat stress exposed chicken was illustrated in table (5)

Feeding of chicken on diet supplemented with *Saccharomyces's* (2.5×10^7 CFU/g) induced significant increase ($P < 0.05$) in total protein & albumin & globulin compared with control group all over the experimental period.

Chicks experimentally exposed to heat stressed and fed on basal diet evoked a significant ($P < 0.05$) decrease in total protein & albumin & globulin all over the experimental period.

Feeding of heat stressed chicken on diet supplemented with *Saccharomyces's* (2.5×10^7 CFU/g) displayed a significant ($P < 0.05$) increase in total protein & albumin & globulin compared with heat stressed group all over the experimental period.

Table 5: The effect of *Saccharomyces's* on protein level (total protein, albumin and globulin) in broiler chicken exposed to heat stress on the 15th and 35st days of age.

Days	15 th			35 st		
	Total protein	Albumin	Globulin	Total protein	Albumin	Globulin

Groups						
Group 1	4.28±0.05b	2.32±0.03bc	1.96±0.04ab	4.58±0.07c	2.44±0.05bc	2.14±0.08cd
Group 2	3.17±0.05c	2.22±0.03cd	0.95±0.06d	3.38±0.03d	2.22±0.03d	1.16±0.04e
Group 3	4.68±0.27a	2.52±0.03a	2.1±0.26bc	4.82±0.03ab	2.48±0.03ab	2.34±0.06cb
Group 4	3.94±0.07b	2.38±0.03b	1.56±0.05c	4.4±0.07c	2.34±0.04bc	2.06±0.08d

Letter differ at $p < 0.05$.

5--Effect of *Saccharomyces's* on glucose & PH

The effect of *Saccharomyces's* on glucose& PH of heat stress exposed chicken was illustrated in table (6)

Feeding of chicken on diet supplemented with *Saccharomyces's* (2.5×10^7 CFU/g) induced non-significant ($P < 0.05$) change on glucose & PH compared with control group all over the experimental period.

Chicks experimentally exposed to heat stressed and fed on basal diet evoked a significant ($P < 0.05$) increase in glucose & PH all over the experimental period.

Feeding of heat stressed chicken on diet supplemented with *Saccharomyces's* (2.5×10^7 CFU/g) displayed a significant ($P < 0.05$) decrease in glucose & PH compared with heat stressed group all over the experimental period.

Table 6: The effect of *Saccharomyces's* on glucose& PH in broiler chicken exposed to heat stress on the 15th and 35st days of age.

Days Groups	15 th		35 st	
	Glucose	PH	Glucose	PH
Group 1	141.4±2.15cd	7.18±0.05d	135.4±2.35d	7.36±0.05d
Group 2	198.6±5.22a	8.392±0.03a	208.8±3.13a	8.38±0.07a
Group 3	140.8±0.73cd	7.232±0.02d	135±3.27d	7.10±0.03e
Group 4	148.2±1.98bc	8.00±0.07c	159.8±2.15b	8.14±0.05b

Letter differ at $p < 0.05$.

6--Effect of *Saccharomyces's* on Haptoglobin & CRP

The effect of *Saccharomyces's* on Haptoglobin & CRP of heat stress exposed chicken was illustrated in table (7)

Feeding of chicken on diet supplemented with *Saccharomyces's* (2.5×10^7 CFU/g) induced significant decrease ($P < 0.05$) on haptoglobin & CRP compared with control group all over the experimental period.

Chicks experimentally exposed to heat stressed and fed on basal diet evoked a significant ($P < 0.05$) increase in haptoglobin & CRP all over the experimental period.

Feeding of heat stressed chicken on diet supplemented with *Saccharomyces's* (2.5×10^7 CFU/g) displayed a significant ($P < 0.05$) decrease in haptoglobin & CRP compared with heat stressed group all over the experimental period.

Table 7: The effect of *Saccharomyces's* on Haptoglobin & CRP in broiler chicken exposed to heat stress on the 15th and 35st days of age.

Days Groups	15 th		35 st	
	Heptoglobin	CRP	Heptoglobin	CRP
Group 1	40.4±0.50bc	4.86±0.47c	135.4±2.35bc	5.86±0.25c
Group 2	53.8±1.42a	13.38±0.66a	208.8±3.13a	17.66±1.29a
Group 3	40±0.74d	3.06±0.19d	135±3.27d	2.6±0.24d
Group 4	42.4±0.50b	6.58±0.18b	159.8±2.15b	8.64±0.40b

Letter differ at $p < 0.05$.

7--Effect of *Saccharomyces's* on oxidant & anti-oxidant stress (MDA & SOD & GPX & catalase & Total antioxidant capacity & Cortisol)

The effect of *Saccharomyces's* on oxidant & anti-oxidant **stress** of heat stress exposed chicken was illustrated in table (8,9,10)

1) MDA

Feeding of chicken on diet supplemented with *Saccharomyces's* (2.5×10^7 CFU/g) induced non-significant ($P < 0.05$) change on MDA compared with control group on 15th days of age and significant ($P < 0.05$) decrease on 35st days of age.

Chicks experimentally exposed to heat stressed and fed on basal diet evoked a significant ($P < 0.05$) increase in MDA all over the experimental period.

Feeding of heat stressed chicken on diet supplemented with *Saccharomyces's* (2.5×10^7 CFU/g) displayed a significant ($P < 0.05$) decrease in MDA compared with heat stressed group all over the experimental period.

2) SOD & GPX & catalase & Total antioxidant capacity (TAC)

Feeding of chicken on diet supplemented with *Saccharomyces's* (2.5×10^7 CFU/g) induced significant ($P < 0.05$) increase on SOD & catalase & GPX & TAC compared with control group all over the experimental period.

Chicks experimentally exposed to heat stressed and fed on basal diet evoked a significant ($P < 0.05$) decrease in SOD & catalase & GPX & TAC all over the experimental period.

Feeding of heat stressed chicken on diet supplemented with *Saccharomyces's* (2.5×10^7 CFU/g) displayed a significant ($P < 0.05$) increase in SOD & catalase & GPX & TAC compared with heat stressed all over the experimental period.

c) Cortisol

Feeding of chicken on diet supplemented with *Saccharomyces*'s (2.5×10^7 CFU/g) induced significant ($P < 0.05$) decrease on cortisol compared with control group all over the experimental period.

Chicks experimentally exposed to heat stressed and fed on basal diet evoked a significant ($P < 0.05$) increase in cortisol all over the experimental period.

Feeding of heat stressed chicken on diet supplemented with *Saccharomyces*'s (2.5×10^7 CFU/g) displayed a significant ($P < 0.05$) decrease in cortisol compared with heat stressed group all over the experimental period.

Table 8: The effect of *Saccharomyces*'s on MDA& SOD in broiler chicken exposed to heat stress on the 15th and 35st days of age.

Days Groups	15 th		35 st	
	MDA	SOD	MDA	SOD
Group 1	14.2±1.24cd	2.94±0.18f	19.6±0.67bc	3.62±0.14e
Group 2	24.4±1.68a	2.5±0.20d	27.00±2.23a	1.16±0.05f
Group 3	12.2±0.58cd	5.78±0.17c	15.6±1.50d	6.38±0.05c
Group 4	19.2±0.58b	4.48±0.24e	20.2±0.37bc	3.24±0.14e

Letter differ at $p < 0.05$.

Table 9: The effect of *Saccharomyces*'s on catalase& GPX in broiler chicken exposed to heat stress on the 15th and 35st days of age.

Days Groups	15 th		35 st	
	CAT	CPX	CAT	CPX
Group 1	3.56±0.16e	21.00±0.83cd	3.9±0.12d	20.80±0.73ef
Group 2	2.6±0.17f	6.40±0.67f	2.04±0.05f	4.60±0.60g
Group 3	6.02±0.32b	28.8±0.86b	6.66±0.22b	32.60±1.12c
Group 4	4.00±0.07de	14.40±1.02e	3.48±0.08e	19.40±0.50f

Letter differ at $p < 0.05$.

Table 10: The effect of *Saccharomyces*'s on total antioxidant capacity& cortisol in broiler chicken exposed to heat stress on the 15th and 35st days of age.

Days Groups	15 th		35 st	
	TAC	Cortisol	TAC	Cortisol
Group 1	250.8±19.40cd	11.6±0.50cd	255.8±7.15d	12.00±0.70b
Group 2	191.4±5.30e	17.6±0.50a	168.4±3.90f	20.4±0.67a
Group 3	301.8±4.31b	9.2±0.66e	327.6±4.98c	8.00±0.44e
Group 4	224.8±3.05d	13.6±0.50b	230.4±3.12e	12.6±0.40b

Letter differ at $p < 0.05$.

8--Effect of *Saccharomyces*'s on Antibody titer by ELISA against ND & IB

The effect of *Saccharomyces*'s on Antibody titer against ND & IB of heat stress exposed chicken was illustrated in table (11)

Feeding of chicken on diet supplemented with *Saccharomyces*'s (2.5×10^7 CFU/g) induced significant ($P < 0.05$) increase on antibody titer of ND & IB compared with control group.

Chicks experimentally exposed to heat stressed and fed on basal diet evoked a significant ($P < 0.05$) decrease on antibody titer of ND & IB compared with control group.

Feeding of heat stressed chicken on diet supplemented with *Saccharomyces*'s (2.5×10^7 CFU/g) displayed a significant ($P < 0.05$) increase on antibody titer of ND & IB compared with heat stressed group.

Table 11: The effect of *Saccharomyces*'s on antibody titer estimated by ELISA against ND& IB in broiler chicken exposed to heat stress.

Groups	ELISA ND	ELISA IB
Group 1	4.14±0.05 ^d	3.6±0.11 ^d
Group 2	2.2±0.07 ^f	2.44±0.05 ^g
Group 3	5.86±0.21 ^b	4.24±0.05 ^c
Group 6	3.34±0.05 ^e	2.94±0.05 ^f

Letter differ at $p < 0.05$.

9- Histopathological investigation:

The effect of *Saccharomyces*'s on tissue histology (Liver- Heart- Lung- Intestine) of heat stress exposed chicken was illustrated in table (12)

table (12):

Groups	Liver	Heart	Lung	Intestine
Group (1)	normal tissue histology	normal tissue histology	normal tissue histology	normal tissue histology
Group (2)	1) severe degree of congestion of blood vessels 2) perivascular edema 3) degeneration of hepatic cells 4) multiple necrotic foci replaced with leukocytes' aggregation	1) severe degree of congestion 2) thickening of blood vessels wall 3) perivascular edema. 4) Degeneration of cardiac myofibril besides myocarditis and leukocytes' infiltration	1) severe degree of congestion of blood vessels with perivascular edema 2)diffuse interstitial hemorrhages 3)Pneumonia and bronchiolitis	1) severe degree of degeneration 2) desquamation of lining epithelium. 3) Congestion and thickening of blood vessels wall in lamina propria
Group (3)	normal tissue histology	normal tissue histology	normal tissue histology	normal tissue histology
Group (4)	revealed milder degree of the mentioned lesions in group 2.	moderate degree of congestion, edema	moderate degree of congestion, edema and inflammation.	slight elongation of length of villi.

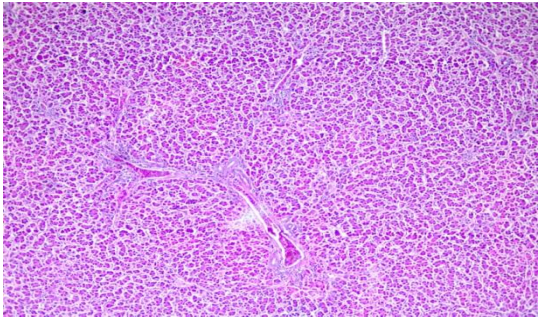


Figure (1): Liver of control negative group showing apparently normal hepatic cell HE, X100.

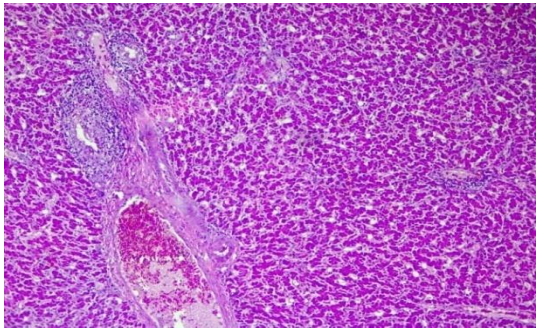


Figure (2): liver of heat stressed group showing congested blood vessels and sinusoids with few perivascular leukocytic infiltraion.HE,X100.

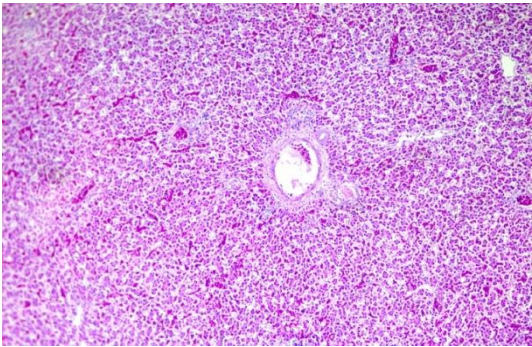


Figure (3): liver of supplemented heat stressed group by Saccharomyces showing mild congested blood vessels and sinusoids .HE,X100.

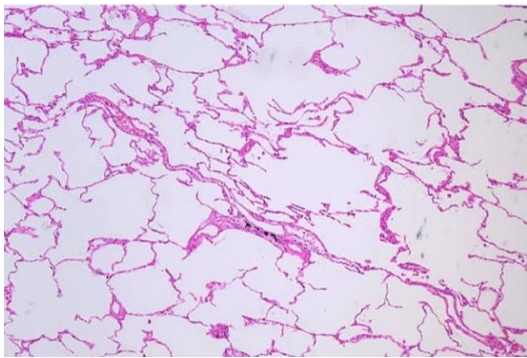


Figure (4): Lung of control negative group showing apparently normal alveoli HE, X100.

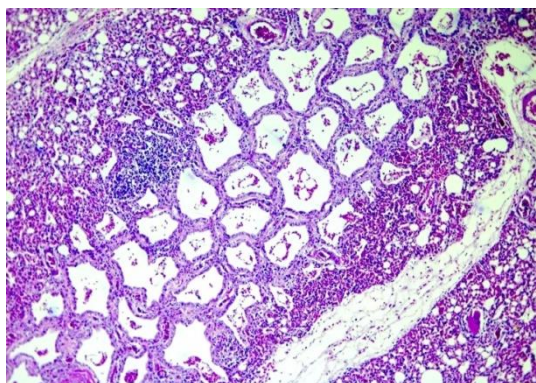


Figure (5): lung of heat stressed group, showing thickening of interalveolar septa , accumulation of eosinophilic material inside the alveoli, hemorrhages and leukocytic agrrgation.HE,X100.

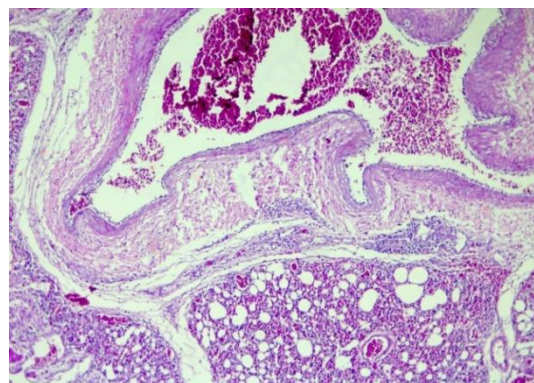


Figure (6):lung of supplemented heat stressed group by Saccharomyces, showing low degree of bronchiolitis and low leukocytes' aggregation HE, X100.

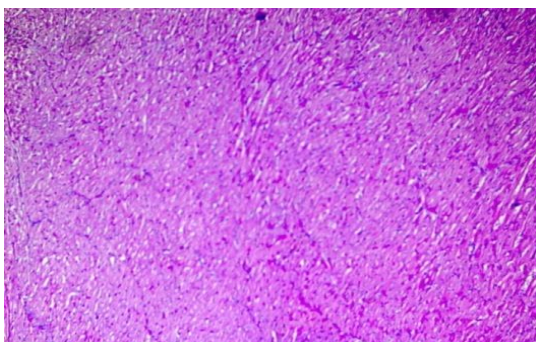


Figure (7): heart of control negative group showing apparently normal tissue HE, X100.

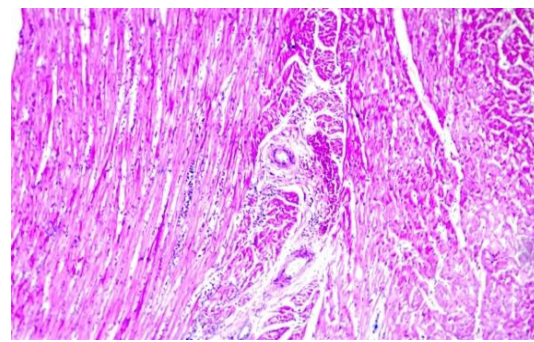


Figure (8): heart of heat stressed group showing hemorrhage and leukocytes' infiltration HE, X100.

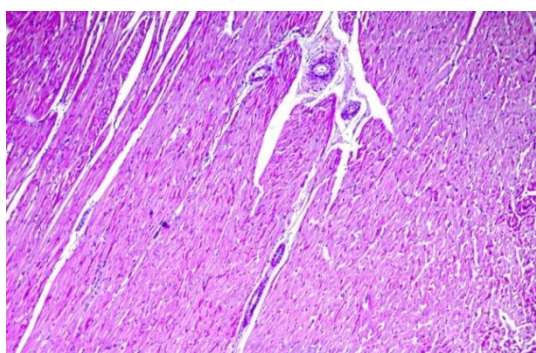


Figure (9):heart of supplemented heat stressed group by Saccharomyces showing mild perivascular edema and few leukocytes' infiltration HE, X100.

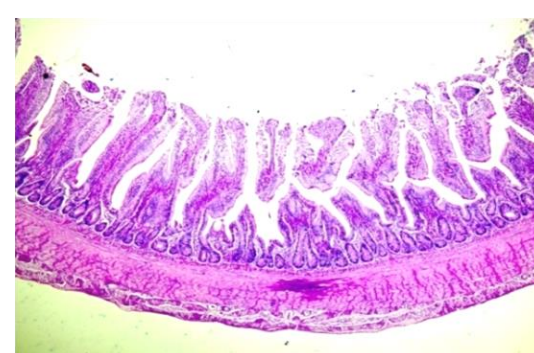


Figure (10): intestine of control negative group showing nearly normal histology HE, X50.

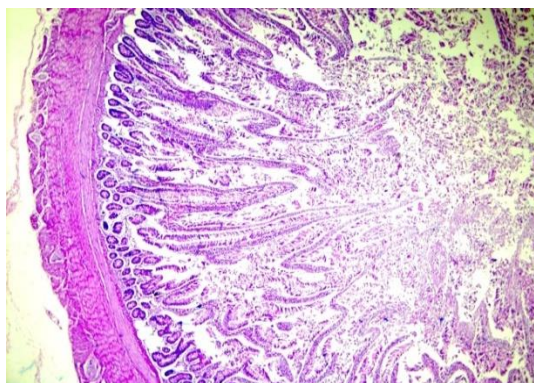


Figure (11):intestine of heat stressed group showing degenerated and sloughed epithelial lining HE, X50.

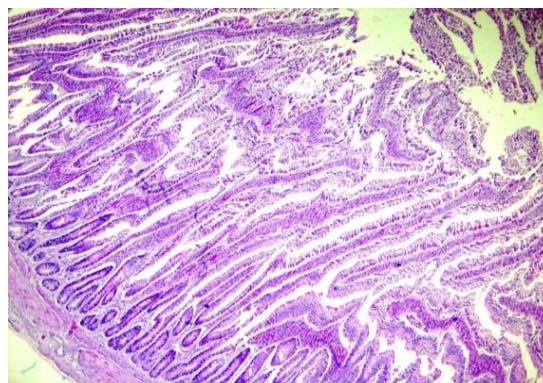


Figure (12):intestine of supplemented heat stressed group by *Saccharomyces*, showing long villi with few degenerated and sloughed epithelial lining. HE, X50.

Discussion

The effect on growth performance parameters

One of the most serious climate problems of tropical and subtropical regions of world negatively affects the production performance of poultry and livestock. Concisely, HS is characterized by endocrine disorders, reduced metabolic rate, lipid peroxidation, decreased feed consumption, higher feed conversion ratio (FCR), immunosuppression, and intestinal microbial dysbiosis (Ashraf et al., 2013; Sohail et al., 2012).

Heat stress results from a negative balance between the net amounts of energy flowing from the animal's body to its surrounding environment and the amount of heat energy produced by the animal. This imbalance may be caused by variations of a combination of environmental factors (e.g: sunlight, thermal irradiation, and air temperature, humidity and movement) and characteristics of the animal (e.g: species, metabolism rate and thermoregulatory mechanisms). Environmental stressors, such as heat stress, are particularly detrimental to animal agriculture (Nienaber et al., 2007–Renaudeau et al., 2012).

In the present study, attempts were made to evaluate the use of *Saccharomyces*'s to determine the effect of such feed supplement on growth performance, some biochemical parameters, oxidative stress, antioxidant status, and histopathological of *heat stressed* chickens.

In broiler nutrition, probiotic species have a beneficial effect on broiler performance (Kalavathy et al., 2003 and Gil De Los Santos et al., 2005).

In present study, a significant increase in body weight was recorded on 15th and 35st days of age in chicken supplemented with probiotics. This may be attributed to the optimum digestion of the feed due to the secretion of microbial digestible enzyme in these chicken and activation of normal flora by creation a favorable media for its growth and regeneration of intestinal villi enhancing nutrient absorption (Kabir et al., 2004).

Probiotic have been administered to counteract such stresses which subsequently improve body weight gain, food consumption in broiler chickens (Hooge et al., 2004). Results of the present study showed that the inclusion of *S. cerevisiae* yeast in broilers ration improved body weight gain, feed intake and food consumption. The obtained results confirmed the previous findings of several researchers (Peralta et al., 2018). Also in agreement with our study, Onifade et al. (1999) reported that SC improved feed/gain ratio and BW gain.

Castro et al., 2012 reported that feed/gain ratio of broiler chicks from 0 to 9 wk of age improved significantly as the SC level in the diets increased.

Several studies gave evidence that the addition of probiotics to the diets of broilers leads to improved performance (**Khaksefidi and Ghooarchi, 2006**).

heat stress constitutes a significant problem leading to a serious economic loss because of its dangerous impacts on health and production (**Wasti et al., 2021**).

In the current study, we found that Heat Stress significantly decreased body weight, body weight gain, food consumption and significantly increased FCR and mortality in broiler chickens. It is reported that HS induces the secretion of stress hormones, which alter the chickens' neuroendocrine system by activating the hypothalamic-pituitary-adrenal axis and thereby increasing the plasma corticosterone levels (**Quinteiro-Filho et al., 2012**). Corticosterone is associated with a higher degree of body protein breakdown (**Yunianto et al., 1997**), which affects the digestive system, nutrient utilization, and digestibility (**Olfati et al., 2018**). Furthermore, HS has been reported to disturb intestinal barrier function, lead to inflammatory responses, and compromise performance. A study by **Alhenaky et al. (2017)** showed that HS, whether chronic or acute, impaired intestinal integrity and increased intestinal permeability to endotoxins and *Salmonella* spp.

The dangerous impacts of HS on broilers growth performance have been recorded by (**Akhavan-Salamat and Ghasemi, 2015**). The decrease in performances of poultry exposed to HS have been accompanied with many reasons as poor appetite and lowered feed intake as a way to reduce heat increase (**Sohail et al., 2013**), badly affected digestion as a result of intestinal morphology damage and decreased activity of digestive enzyme (**Chen et al., 2014**) and impaired metabolism owing to reduced activity of thyroid hormones (**Sohail et al., 2010**), altered status of endocrine as increased level of corticosterone hormone (**Sohail et al., 2012**).

The positive impact of SC probiotic on broilers reared under normal and HS conditions may be due to offering competitive binding sites for harmful bacteria, as prebiotic microorganisms have been shown to maintain the expansion of undesirable intestinal pathogenic microorganisms in broiler birds (**Tian et al., 2016**). This activity has been shown to enhance GIT health state, nutrition absorption, and growth performance by reducing competition between pathogenic bacteria and the host for nutrients (**Spring et al., 2000**). The yeast (SC) possesses a probiotic activity which decreased the infection chances in poultry (**Ghasemi et al., 2006**). The probiotics impact of yeast has been found to enhance feed conversion efficiency (**Ayanwale et al., 2006**), enhance BWG and decrease mortality (**Jin et al., 2004**) and stimulate the immune organs (**Elfaky et al., 2010**). The SC cell wall could improve the intestinal mucosa aspects and this is suggested that it could be the explanation for the improvement in broilers performance provided with SC **Santin et al. (2001 and 2003)**. Concordantly, this was emphasized by **Miazzo et al. (2005)** who stated that the broiler chicks fed on diet supplemented with yeast to replace a part of the premix showed a better average WG and FC ratio. **Yang et al. (2012)** who stated that Mannan oligosaccharide, obtained from the outer cell wall of yeast (*Saccharomyces cerevisiae*), can be used as a growth promoter as well as a possible antibiotics alternative to in broiler rations. **Sohail et al. (2012)** who reported that supplementing *Saccharomyces cerevisiae* probiotic to broilers reared under HS conditions significantly improved growth performance which is consistent with the findings of this experiment. Recently (**Ahiwe et al., 2020**) who stated that yeast as a whole and its cell walls dietary supplementation in broilers at 1.5–2 g/kg level could enhance growth performance and meat yield.

The effect on Liver function (ALT &AST)

Measurement of serum transaminase ALT, AST activities are standard test for hepatocellular damage. It's well known that the enzyme are intracellular substances being located in the mitochondria and the cytoplasm, consequently, circulating levels increase following liver cell damage. (**Doxey, 1971**).

Regarding the biochemical findings, HS significantly ($P < 0.05$) elevated serum AST and ALT activities levels when compared to NHS birds group. Increased serum AST and ALT levels can take place as a result of alteration of hepatocellular membrane either owing to blood hypoxia, toxins and toxemia exposure, metabolic

disorders inflammation, or hepatic cells proliferation. Increased AST and ALT activities were also recorded with the damage of liver and intestine (**Rani et al., 2011**).

These findings seemed compatible with (**Lu-PingTang et al., 2022**) who reported that broiler chicks exposed to heat stress for 1 or 2 weeks displayed necrotic points on hepatic surface. Moreover, hepatic histological examination provoked inflammatory cells infiltration. These results indicated that heat stress induced hepatic tissue damage.

In the current study the decrease in serum AST and ALT levels by SC supplemented groups was recorded it may collectively indicate the normal functions of liver and intestine of chickens fed feeds containing SC during HS conditions. This result is also confirmed by the obtained results of **Olugbemi et al. (2010)** who stated that SC possess a beneficial action to stimulate the immune responses and enhance intestinal health in broilers. Moreover, this could give a reflection about the protective effect of SC addition on hepatic tissues.

The effect on Kidney function (creatinine & uric acid)

Regarding the kidney function, several changes were recorded uric acid is the primary catabolic product of protein catabolism in birds (**Harrison and Harrison, 1986**). The avian kidney excretes uric acid primarily by tubular excretion, unlike the mammalian system that excretes urea entirely by filtration therefore the elevation of serum uric acid level indicate impaired renal function (**Osbaldiston, 1968**).

In the current study heat stress significantly ($P < 0.05$) increased serum creatinine and uric acid levels as matched to NHS group.

Serum uric acid and creatinine levels measurement is the most sensitive indicators to evaluate renal functions and it is often considered an important tool in the evaluation of the Kidney state. In the current study, the obtained results seemed compatible to that recorded by **Tang et al. (2018)** who stated that birds subjected to HS displayed increased urea and uric acid levels in plasma, which indicated renal dysfunction.

Huang et al. (2018) who stated that acute HS caused an increase in some hemato biochemical parameters level such as creatinine, blood urea nitrogen, ALT and creatine kinase that may referee to the injury of some critical organs like kidney and liver.

Huff et al. (2013) recorded that the dietary inclusion of SC in broilers ration, significantly ($P < 0.05$) reduced uric acid and creatinine.

Effect on protein level

Serum protein is the total amount of protein in the blood in which albumin is the most abundant. Albumin helps keep the blood from leaking out of blood vessels as well as to carry some medicines and other substances through the blood that is important for tissue growth and healing. Low total protein levels can suggest a liver disorder, a kidney disorder, or a disorder in which protein is not digested or absorbed properly.

Heat stress significantly ($P < 0.05$) decreased serum total protein, albumin and globulin levels as matched to NHS group. In the current study, the obtained results seemed compatible to that recorded by **Sahin (2012)** also reported that heat stress inhibited the levels of protein and albumin and alleviated the effect of heat stress by supplement MOL in diet.

In the present study, SC supplementation adversely influenced serum protein and albumin concentrations, Several authors have identified that probiotic supplementation in diets increased serum protein and albumin concentrations, in broilers (**Agawane and Lonkar, et al., 2004, Capcarova et al., 2011**), piglets (**Harding et al., 2008**) and fish (**Zhou et al., 2010**) reported that probiotics did not stimulate gastrointestinal protein synthesis or reduce the severity of intestinal colitis, but reported an unidentified signaling mechanism between the gut and liver which could be responsible for the probiotic-induced increase in liver protein and plasma protein synthesis. Furthermore, dietary probiotic supplementation may increase the serum protein and albumin concentrations by increasing iron absorption in the lower intestine (**Eizaguirre et al., 2002**).

Effect on Glucose level

In the present study it has been shown that heat stressed chicks demonstrate a significant increase in the level of serum glucose. The increase in glucose concentration is directly responsive to an increase in glucocorticoids (**Borges et al., 2007**), which can result from various stressors including heat stress. Glucocorticoids have primary effects on metabolism, stimulating gluconeogenesis from muscle tissue proteins. **Habibian et al. (2014)** reported that high environmental temperature increased levels of plasma glucose and cholesterol and reduced protein level.

On the contrary, the dietary supplementation of SC in bird ration reared under HS significantly ($P < 0.05$) lowered the serum glucose compared to HS non treated group. These results agree with findings of **Djouvinov et al. (2005)** who tested the probiotic strains in ducklings.

Effect on PH value

HS significantly increased the production of lactic acid thereby fasting the pH decline rate and consequently decreasing the breast muscle quality, which lastly leads to pale, soft, and exudative meat (PSE-like meat) in poultry (**Wang et al., 2017**). HS could induce a sharp decrease in the metabolism of birds, which in turn will produce dangerous complications, such as a color change, decrease in muscle pH and water-holding capacity (WHC) of chicken meat (**Gonzalez-Rivas et al., 2020**).

Cramer et al. (2018) reported that providing with probiotic could mitigate the pH decrease of breast muscles from heated stressed broiler by likely affecting the rate of postmortem glycolysis metabolism.

Effect on Haptoglobin & CRP

Haptoglobin is an alpha-2 globulin that is known as PIT 54 in chickens. Haptoglobin's major role is to bind free hemoglobin produced from erythrocytes and decrease its oxidative activity (**Yang et al., 2003**). It also binds hemoglobin to reduce iron losses via urine after hemolysis, hence sparing tissues from harm caused by free hemoglobin (**Nielsen et al., 2006**). Haptoglobin has been identified as a potent angiogenic agent essential for endothelial cell proliferation and differentiation in the development of new blood vessels.

CRP was the first APP been described and is one of the major APP in humans and dogs (**Eckersall and Bell, 2010**). CRP has the ability to binds directly to degenerating cells, residues and polysaccharides on bacteria, fungi, and parasites, acting as an opsonin. It can activate the complement system when bound to one of its ligands and can bind to phagocytic cells, interacting together with humoral and cellular systems of inflammation (**Cray et al., 2009**).

In the current study the decrease in serum heptoglobin and c- reactive protein levels in SC supplemented groups was recorded it may collectively indicate the normal functions of liver of chickens fed feeds containing SC during HS conditions. This result is also confirmed by the obtained results of **Maoba et al. (2021)**, who stated that SC possess a beneficial action to stimulate the immune responses and anti inflammatory in broilers.

Oxidant & Antioxidant stsatus

Regarding to the antioxidant status, in the present study broilers fed basal diets and raised under HS conditions divulged significant reduction in catalase (CAT), super oxide dismutase (SOD), glutathione peroxidase (Gpx), total antioxidant capacity (TAC) activities together with increased MDA levels and cortisol.

HS is considered one of the most important factors having the ability of enhancing ROS production (**Li et al., 2020**). These phenomena resulted in oxidative stress via affecting function of mitochondria and changing ROS levels, therefore causing oxidative damage of lipids and proteins and change of oxidative stress markers levels, such as MDA, GPx, catalase and SOD (**Emami et al., 2021**).

probiotic, as an antioxidant is able to restore the SOD activity and reduces serum MDA level in broilers raised under HS. This is emphasized by **Bu et al. (2019)** and **Timo thee Andriamialinirina et al. (2020)** who noted that supplementation with products derived from Yeast involving culture of yeast were reported to

improve both serum and liver tissue GPX, SOD, and CAT enzyme activities and reduced MDA level in animals. Carbohydrates in yeast might share a role in enhancing antioxidant efficiency (**Wang et al., 2021**).

Effect on anti body titers against ND & IB

In the current study, mean antibody titer against ND and IB increased significantly in heat stressed group. It has been postulated that heat stress increases the corticosterone level, an immunosuppressive agent, while SC decrease the level of corticosterone (**Rama et al. 2004**).

Heat stress increases the production of free radicals and reduces the concentration of antioxidant vitamins like vitamin C, E, and A and minerals (**Khan et al., 2011**). Findings of the present study are in agreement to that of **Khajavi et al. (2003)**, who worked on chicken under heat-stressed condition and reported significant reduction in antibodies production. **Rama et al. (2004)** reported that heat stress reduces antibody production by increasing secretion of immunosuppressive agents (hydrocortisones), while SC reduces hydrocortisones production. Similarly, **Coelho and McNaughton (1995)** reported that higher vitamin supplementation enhanced broiler performance and immunity in stress condition, which are in agreement with the findings of the present study.

zhang et al. (2005) reported that antibody titers against NDV& IB was improved in broiler chickens fed SC.

Histopathological investigation

Prime examples of heat stress were increased heart rate and increased blood flow to muscle, brain, and heart (**Ewing et al., 1999**). This might cause congestion in kidney, liver and lung. Effects of heat stroke on the gross lesions were dominated by severe and generalized hyperemia, which was most severe in the respiratory tract, especially in the lung, tracheal and bronchial mucosae. Lungs may also be edematous and occasionally contain focal consolidations of bronchopneumonia. Other organs, such as heart, kidney, may also be severely congested. The microscopic lesions are compatible with those seen grossly. The vasculatures of the lung are severely engorged and edema are evident in alveoli. Centritubular necrosis, dissociation of hepatocytes and congestion are often found in the liver. Subendocardium and subepicardial hemorrhages are found in the heart. Besides, capillary congestion is evident in kidney and other structures (**Smith et al., 1979**).

Conclusion

It could be concluded that supplementation of chickens with probiotic (*Saccharomyces*) had a significant effect on growth performance & some biochemical parameters (ALT, AST, Creatinine, Uric acid, total protein, albumin, globulin, glucose, Ph, Haptoglobin, CRP) & oxidant and antioxidant parameter (GSH, SOD, Catalase, Total antioxidant capacity, cortisol) & histopathological condition. Based on the result of present study.

References

1. **Aalaei, M., Khatibjoo, A., Zaghari, M., Taherpour, K., Akbari Gharaei, M., & Soltani, M. (2018):** Comparison of single and multi-strain probiotics effects on broiler breeder performance, egg production, egg quality and hatchability. *British poultry science*, 59(5), 531-538.
2. **Abudabos, A. M., Alyemni, A. H., Dafalla, Y. M., & Khan, R. U. (2016):** The effect of phytogenic feed additives to substitute in-feed antibiotics on growth traits and blood biochemical parameters in broiler chicks challenged with *Salmonella typhimurium*. *Environmental Science and Pollution Research*, 23, 24151-24157.
3. **Agawane, S. B., & Lonkar, P. S. (2004):** Effect of probiotic containing *Saccharomyces boulardii* on experimental ochratoxicosis in broilers: hematobiochemical studies. *Journal of veterinary science*, 5(4), 359-367.
4. **Ahiwe, E. U., Abdallah, M. E., Chang'a, E. P., Omede, A. A., Al-Qahtani, M., Gausi, H., ... & Iji, P. A. (2020):** Influence of dietary supplementation of autolyzed whole yeast and yeast cell wall products on broiler chickens. *Asian-Australasian journal of animal sciences*, 33(4), 579.

5. **Ahmad, I. (2006):** Effect of probiotics on broilers performance. *International Journal of Poultry Science*, 5(6), 593-597.
6. **Akhavan-Salamat, H., & Ghasemi, H. A. (2015):** Alleviation of chronic heat stress in broilers by dietary supplementation of betaine and turmeric rhizome powder: dynamics of performance, leukocyte profile, humoral immunity, and antioxidant status. *Tropical animal health and production*, 48, 181-188.
7. **Alhenaky, A., Abdelqader, A., Abuajamieh, M., & Al-Fataftah, A. R. (2017):** The effect of heat stress on intestinal integrity and Salmonella invasion in broiler birds. *Journal of thermal biology*, 70, 9-14.
8. **Ashraf, S., Zaneb, H., Yousaf, M. S., Ijaz, A., Sohail, M. U., Muti, S., ... & Rehman, H. (2013):** Effect of dietary supplementation of prebiotics and probiotics on intestinal microarchitecture in broilers reared under cyclic heat stress. *Journal of animal physiology and animal nutrition*, 97, 68-73.
9. **Ayanwale, B. A., Kpe, M., & Ayanwale, V. A. (2006):** The effect of supplementing *Saccharomyces cerevisiae* in the diets on egg laying and egg quality characteristics of pullets. *International journal of poultry science*, 5(8), 759-763.
10. **Borges, S. A., Da Silva, A. F., & Maiorka, A. (2007):** Acid-base balance in broilers. *World's Poultry Science Journal*, 63(1), 73-81.
11. **Botsoglou E, Govarisi A, Moulas A, Botsoglou N (2010):** Oxidative stability and microbial growth of turkey breast fillets during refrigerated storage as influenced by feed supplementation with olive leaves, oregano and/or α -tocopheryl acetate. *Br. Poult. Sci.* 51:760-768.
12. **Bu XY, Lian XQ, Wang Y, Luo CZ, Tao SQ, Liao YL, Yang JM, Chen AJ, Yang YH (2019):** Dietary yeast culture modulates immune response related to TLR2-MyD88-NF- κ B signaling pathway, antioxidant capability and disease resistance against *Aeromonas hydrophila* for Ussuri catfish (*Pseudobagrus ussuriensis*) *Fish Shellfish Immunol.*; 84:711–718.
13. **Capcarova, M., Hascik, P., Kolesarova, A., Kacaniova, M., Mihok, M., & Pal, G. (2011):** The effect of selected microbial strains on internal milieu of broiler chickens after peroral administration. *Research in Veterinary Science*, 91(1), 132-137.
14. **Castro, M., & Corredor, G. A. (2012):** The effect of dietary inclusion of commercial yeast (*Saccharomyces cerevisiae*) on productive parameters of broilers. An experience in the classroom. *Revista de Investigaciones de UNIAGRARIA*, 1(1), 23-34.
15. **Chen, P., Zhang, Q., Dang, H., Liu, X., Tian, F., Zhao, J., ... & Chen, W. (2014):** Screening for potential new probiotic based on probiotic properties and α -glucosidase inhibitory activity. *Food Control*, 35(1), 65-72.
16. **Coelho, M. B., & McNaughton, J. L. (1995):** Effect of composite vitamin supplementation on broilers. *Journal of Applied Poultry Research*, 4(3), 219-229.
17. **Cramer T, Kim H, Chao Y, Wang W, Cheng H, Kim Y (2018):** Effects of probiotic (*Bacillus subtilis*) supplementation on meat quality characteristics of breast muscle from broilers exposed to chronic heat stress. *Poultry science*, 97: 3358-3368.
18. **Cray, C., Zaias, J., & Altman, N. H. (2009):** Acute phase response in animals: a review. *Comparative medicine*, 59(6), 517-526.
19. **Demeke, S. (2004):** Growth performance and survival of local and white Leghorn chicken under intensive management system. *SINET: Ethiopian Journal of Science*, 27(2), 161-164.
20. **Djouvinov, D., Stefanov, M., Boicheva, S., & Vlaikova, T. (2005):** Effect of diet formulation on basis of digestible amino acids and supplementation of probiotic on performance of broiler chicks. *Trakia J. Sci*, 3, 61-69.
21. **Doxey, D. L. (1971):** *Veterinary clinical pathology* (pp. x+-356).
22. **Dugas, B., Mercenier, A., Lenoir-Wijnkoop, I., Arnaud, C., Dugas, N., & Postaire, E. (1999):** Immunity and probiotics. *Immunology today*, 20(9), 387-390.
23. **Eckersall, P. D., & Bell, R. (2010):** Acute phase proteins: Biomarkers of infection and inflammation in veterinary medicine. *The veterinary journal*, 185(1), 23-27.

24. **Edens, F. W. (2003):** An alternative for antibiotic se in poultry: probiotics. *Brazilian Journal of Poultry Science*, 5, 75-97.
25. **Eizaguirre, I., Aldazabal, P., Urkia, N. G., Asensio, A., & Arenzxana, J. M. G. (2011):** Escherichia coli translocation in experimental short bowel syndrome: probiotic supplementation and detection by polymerase chain reaction. *Pediatric surgery international*, 27, 1301-1305.
26. **Elfaky, O. A. (2010):** Effect of Dietary Supplementation of Yeast (*Saccharomyces cerevisiae*) on Performance and Carcass Characteristics of Broiler Chicks. *Animal Production University of Gezira*.
27. **Elghandour, M. M. Y., Tan, Z. L., Abu Hafsa, S. H., Adegbeye, M. J., Greiner, R., Ugbogu, E. A., ... & Salem, A. Z. M. (2020):** *Saccharomyces cerevisiae* as a probiotic feed additive to non and pseudo-ruminant feeding: a review. *Journal of applied microbiology*, 128(3), 658-674.
28. **El-Kholy MS, El-Hindawy MM, Alagawany M, Abd El-Hack ME, Sabry AAE (2018):** Use of acetylsalicylic acid as an allostatic modulator in the diets of growing Japanese quails exposed to heat stress. *J. Therm. Biol.* 74, 6–13.
29. **Emami NK, Jung, U Voy B and Dridi S (2021):** “Radical response: effects of heat stress-induced oxidative stress on lipid metabolism in the avian liver,” *Antioxidants*, vol. 10, no. 1, p. 35.
30. **Ewing, R. C. (1999):** Nuclear waste forms for actinides. *Proceedings of the National Academy of Sciences*, 96(7), 3432-3439.
31. **Felver-Gant, J. N., Dennis, R. L., Zhao, J., & Cheng, H. W. (2014):** Effects of dietary antioxidant on performance and physiological responses following heat stress in laying hens. *International Journal of Poultry Science*, 13(5), 260.
32. **Ghasemi, H. A., Tahmasbi, A. M., Moghaddam, G. H., Mehri, M., Alijani, S., Kashefi, E., & Fasihi, A. (2006):** The effect of phytase and *Saccharomyces cerevisiae* (sc47) supplementation on performance, serum parameters, phosphorous and calcium retention of broiler chickens. *International Journal of Poultry Science*, 5(2), 162-168.
33. **Gil De Los Santos, J. R., Storch, O. B., & Gil-Turnes, C. (2005):** *Bacillus cereus* var. *toyoii* and *Saccharomyces boulardii* increased feed efficiency in broilers infected with *Salmonella enteritidis*. *British poultry science*, 46(4), 494-497.
34. **Gonzalez-Rivas, P. A., Chauhan, S. S., Ha, M., Fegan, N., Dunshea, F. R., & Warner, R. D. (2020):** Effects of heat stress on animal physiology, metabolism, and meat quality: A review. *Meat science*, 162, 108025.
35. **Gunal, M., Yayli, G., Kaya, O., Karahan, N., & Sulak, O. (2006):** The effects of antibiotic growth promoter, probiotic or organic acid supplementation on performance, intestinal microflora and tissue of broilers. *International Journal of Poultry Science*, 5(2), 149-155.
36. **Habibian, M., Ghazi, S., Moeini, M. M., & Abdolmohammadi, A. (2014):** Effects of dietary selenium and vitamin E on immune response and biological blood parameters of broilers reared under thermoneutral or heat stress conditions. *International journal of biometeorology*, 58, 741-752.
37. **Harding, S. V., Fraser, K. G., & Wykes, L. J. (2008):** Probiotics stimulate liver and plasma protein synthesis in piglets with dextran sulfate-induced colitis and macronutrient restriction. *The Journal of nutrition*, 138(11), 2129-2135.
38. **Harrison G . J. and Harrison L.R. (1986):** *Clinical Avian Medicine and Surgery*, WB Saunders company Philadelphia, London.Toronto.
39. **Hooge, D. M., Ishimaru, H., & Sims, M. D. (2004):** Influence of dietary *Bacillus subtilis* C-3102 spores on live performance of broiler chickens in four controlled pen trials. *Journal of Applied Poultry Research*, 13(2), 222-228.
40. **Huang S, Yang H, Rehman MU, Tong Z (2018):** Acute heat stress in broiler chickens and its impact on serum biochemical and electrolyte parameters. *Indian Journal of Animal Research* 52:683-686.
41. **Huang, Y., & Adams, M. C. (2004):** In vitro assessment of the upper gastrointestinal tolerance of potential probiotic dairy propionibacteria. *International journal of food microbiology*, 91(3), 253-260.

42. **Huff, G. R., Huff, W. E., Jalukar, S., Oppy, J., Rath, N. C., & Packialakshmi, B. (2013):** The effects of yeast feed supplementation on turkey performance and pathogen colonization in a transport stress/*Escherichia coli* challenge. *Poultry science*, 92(3), 655-662.
43. **Jin, Y. S., Laplaza, J. M., & Jeffries, T. W. (2004):** *Saccharomyces cerevisiae* engineered for xylose metabolism exhibits a respiratory response. *Applied and environmental microbiology*, 70(11), 6816-6825.
44. **Kabir, S. L., Rahman, M. M., Rahman, M. B., Rahman, M. M., & Ahmed, S. U. (2004):** The dynamics of probiotics on growth performance and immune response in broilers. *Int. J. Poult. Sci*, 3(5), 361-364.
45. **Kalavathy, R., Abdullah, N., Jalaludin, S., & Ho, Y. W. (2003):** Effects of *Lactobacillus* cultures on growth performance, abdominal fat deposition, serum lipids and weight of organs of broiler chickens. *British poultry science*, 44(1), 139-144.
46. **Khaksefidi, A., & Ghoorchi, T. (2006):** Effect of probiotic on performance and immunocompetence in broiler chicks. *The Journal of Poultry Science*, 43(3), 296-300.
47. **Khan, R. U., Naz, S., Nikousefat, Z., Tufarelli, V., Javdani, M., Rana, N., & Laudadio, V. (2011):** Effect of vitamin E in heat-stressed poultry. *World's poultry science journal*, 67(3), 469-478.
48. **Li, Q., Wan, G., Peng, C., Xu, L., Yu, Y., Li, L., & Li, G. (2020):** Effect of probiotic supplementation on growth performance, intestinal morphology, barrier integrity, and inflammatory response in broilers subjected to cyclic heat stress. *Animal science journal*, 91(1), e13433.
49. **Lu-PingTang, Yi-Lei Liu, Jia-Xin Zhang, Kang-Ning Ding, Meng-Han Lu and Yong-Ming He (2022):** Heat stress in broilers of liver injury effects of heat stress on oxidative stress and autophagy in liver of broilers. *Poultry Science*, 101(10): 102085.
50. **Maoba, S., Ogbuewu, I. P., Oguttu, J. W., & Mbajiorgu, C. A. (2021):** Haematological profiles of indigenous Boschveld chickens on probiotic-yeast (*Saccharomyces cerevisiae*) supplementation. *Comparative Clinical Pathology*, 30, 293-299.
51. **Miazzo, R. D., Peralta, M. F., Picco, M. L., & Nilson, A. J. (2005):** Productive parameters and carcass quality of broiler chickens fed yeast (*S. cerevisiae*).
52. **Mottet, A., & Tempio, G. (2017):** Global poultry production: current state and future outlook and challenges. *World's poultry science journal*, 73(2), 245-256.
53. **Nielsen, M. J., Petersen, S. V., Jacobsen, C., Oxvig, C., Rees, D., Møller, H. J., & Moestrup, S. K. (2006):** Haptoglobin-related protein is a high-affinity hemoglobin-binding plasma protein. *Blood*, 108(8), 2846-2849.
54. **Nienaber, J. A., Hahn, G. L., Brown-Brandl, T. M., & Eigenberg, R. A. (2007):** Summer heat waves—extreme years. In *2007 ASAE Annual Meeting* (p. 1). American Society of Agricultural and Biological Engineers.
55. **NRC (1994):** Nutrient Requirements of Poultry. Washington DC, USA: National Academy Press.
56. **Olfati, A., Mojtahedin, A., Sadeghi, T., Akbari, M., & Martínez-Pastor, F. (2018):** Comparison of growth performance and immune responses of broiler chicks reared under heat stress, cold stress and thermoneutral conditions. *Spanish Journal of Agricultural Research*, 16(2), e0505-e0505.
57. **Olugbemi, T. S., Mutayoba, S. K., & Lekule, F. P. (2010):** Effect of *saccharomyces* inclusion in cassava based diets fed to broiler chickens. *International Journal of Poultry Science*, 9(4), 363-367.
58. **Onifade, A. A., Obiyan, R. I., Onipede, E., Adejumo, D. O., Abu, O. A., & Babatunde, G. M. (1999):** Assessment of the effects of supplementing rabbit diets with a culture of *Saccharomyces cerevisiae* using growth performance, blood composition and clinical enzyme activities. *Animal Feed Science and Technology*, 77(1-2), 25-32.
59. **Osbaldiston, G. W. (1968):** The effect of climate on the growth performance of populations of broiler chickens. *British Veterinary Journal*, 124(2), 56-68.
60. **Peralta, M. F., Nilson, A. J., & Miazzo, R. D. (2018):** Effect of *Saccharomyces cerevisiae* association with threonine on productive performance in broilers. *Iranian Journal of Applied Animal Science*, 8(4), 677-684.

61. **Quinteiro-Filho, W. M., Gomes, A. V. S., Pinheiro, M. L., Ribeiro, A., Ferraz-de-Paula, V., Astolfi-Ferreira, C. S., ... & Palermo-Neto, J. (2012):** Heat stress impairs performance and induces intestinal inflammation in broiler chickens infected with *Salmonella* Enteritidis. *Avian Pathology*, 41(5), 421-427.
62. **Rama, I., Cruzado, J. M., Gil-Vernet, S., Torras, J., Serón, D., Castela, A. M., ... & Grinyó, J. M. (2005):** Steroids can be safely withdrawn from cyclosporine and mycophenolate mofetil-treated renal allograft recipients: long-term results. *Transplantation*, 80(2), 164-168.
63. **Rani MP, Ahmad, NN, Prasad PE and Latha CS (2011):** Haematological and biochemical changes of stunting syndrome in broiler chicken. *Vet. World*, 4: 124-125.
64. **Renaudeau, D., Collin, A., Yahav, S., De Basilio, V., Gourdine, J. L., & Collier, R. J. (2012):** Adaptation to hot climate and strategies to alleviate heat stress in livestock production. *Animal*, 6(5), 707-728.
65. **Sahin, K., Orhan, C., Tuzcu, Z., Tuzcu, M., & Sahin, N. (2012):** Curcumin ameliorates heat stress via inhibition of oxidative stress and modulation of Nrf2/HO-1 pathway in quail. *Food and Chemical Toxicology*, 50(11), 4035-4041.
66. **Santin, E., Paulillo, A. C., Maiorka, A., Nakaghi, L. S., & Macari, M. (2003):** Evaluation of the efficacy of *Saccharomyces cerevisiae* cell wall to ameliorate the. *Int J Poultry Sci*, 2(5), 341-344.
67. **Smith, L. L., Rose, M. S., & Wyatt, I. (1979, January):** The pathology and biochemistry of paraquat. In *Ciba Foundation Symposium 65-Oxygen Free Radicals and Tissue Damage* (Vol. 65, pp. 321-341). Chichester, UK: John Wiley & Sons, Ltd..
68. **Sohail, M. U., Hume, M. E., Byrd, J. A., Nisbet, D. J., Ijaz, A., Sohail, A., ... & Rehman, H. (2011):** Effect of supplementation of prebiotic mannan-oligosaccharides and probiotic mixture on growth performance of broilers subjected to chronic heat stress. *Poultry science*, 91(9), 2235-2240.
69. **Sohail, M. U., Hume, M. E., Byrd, J. A., Nisbet, D. J., Ijaz, A., Sohail, A., ... & Rehman, H. (2012):** Effect of supplementation of prebiotic mannan-oligosaccharides and probiotic mixture on growth performance of broilers subjected to chronic heat stress. *Poultry science*, 91(9), 2235-2240.
70. **Sohail, M. U., Hume, M. E., Byrd, J. A., Nisbet, D. J., Ijaz, A., Sohail, A., ... & Rehman, H. (2012):** Effect of supplementation of prebiotic mannan-oligosaccharides and probiotic mixture on growth performance of broilers subjected to chronic heat stress. *Poultry science*, 91(9), 2235-2240.
71. **Sohail, M. U., Hume, M. E., Byrd, J. A., Nisbet, D. J., Ijaz, A., Sohail, A., ... & Rehman, H. (2012):** Effect of supplementation of prebiotic mannan-oligosaccharides and probiotic mixture on growth performance of broilers subjected to chronic heat stress. *Poultry science*, 91(9), 2235-2240.
72. **Sohail, M. U., Ijaz, A., Younus, M., Shabbir, M. Z., Kamran, Z., Ahmad, S., ... & Rehman, H. (2013):** Effect of supplementation of mannan oligosaccharide and probiotic on growth performance, relative weights of viscera, and population of selected intestinal bacteria in cyclic heat-stressed broilers. *Journal of Applied Poultry Research*, 22(3), 485-491.
73. **Sohail, M. U., Ijaz, A., Yousaf, M. S., Ashraf, K., Zaneb, H., Aleem, M., & Rehman, H. (2010):** Alleviation of cyclic heat stress in broilers by dietary supplementation of mannan-oligosaccharide and Lactobacillus-based probiotic: Dynamics of cortisol, thyroid hormones, cholesterol, C-reactive protein, and humoral immunity. *Poultry science*, 89(9), 1934-1938.
74. **Spring P, Wenk C, Dawson, KA and Newman, KE (2000):** The effects of dietary mannanoligosaccharides on cecal parameters and the concentrations of enteric bacteria in the ceca of salmonella-challenged broiler chicks *Poult. Sci*, 79, pp. 205-211.
75. **Tang S, Zhou S, Bin Yin B, Xu J, Di L, Zhang J and Bao E (2018):** Heat stress-induced renal damage in poultry and the protective effects of HSP60 and HSP47Cell Stress and Chaperones, 23:1033–1040.
76. **Tian X, Shao Y, Wang Z and GuoY (2016):** Effects of dietary yeast beta-glucans supplementation on growth performance, gut morphology, intestinal *Clostridium perfringens* population and immune response of broiler chickens challenged with necrotic enteritis *Anim. Feed Sci. Technol*, 215, pp. 144-155.

77. **Timothée Andriamialinirina HJ, Irm M, Taj, S, Lou JH, Jin M, Zhou Q (2020):** The effects of dietary yeast hydrolysate on growth, hematology, antioxidant enzyme activities and non-specific immunity of juvenile Nile tilapia, *Oreochromis niloticus*. *Fish Shellfish Immunol*, 101, pp. 168-175.
78. **Wang RH, Liang RR, Lin H, Zhu LX, Zhang YM, Mao YW....and Luo X (2017):** Effect of acute heat stress and slaughter processing on poultry meat quality and postmortem carbohydrate metabolism. *Poultry science*, 96(3), 738-746.
79. **Wang T, Cheng K, Yu CY, Li QM, Tong YC, Wang C, Yang ZB, Wang T (2021):** Effects of a yeast-derived product on growth performance, antioxidant capacity, and immune function of broilers. *Poult Sci.*, 100(9):101343.
80. **Wasti S, Sah N, Singh AK, Lee CN, Jha R, Mishra B (2021):** Dietary supplementation of dried plum: a novel strategy to mitigate heat stress in broiler chickens. *J Anim Sci Biotechnol*. Mar 30;12(1):58. doi: 10.1186/s40104-021-00571-5. PMID: 33781340; PMCID: PMC8008564.
81. **Yang Y, Iji PA and Choct M (2009):** Effects of different dietary levels of mannanoligosaccharide on growth performance and gut development of broiler chickens. *Asian Australas J Anim Sci*. 20:1084–1091.
82. **Yang, C. M., Cao, G. T., Ferket, P. R., Liu, T. T., Zhou, L., Zhang, L., ... & Chen, A. G. (2012):** Effects of probiotic, *Clostridium butyricum*, on growth performance, immune function, and cecal microflora in broiler chickens. *Poultry science*, 91(9), 2121-2129.
83. **Yang, Y., Sharma, R., Sharma, A., Awasthi, S., & Awasthi, Y. (2003):** Lipid peroxidation and cell cycle signaling: 4-hydroxynonenal, a key molecule in stress mediated signaling. *Acta Biochimica Polonica*, 50(2), 319-336.
84. **Yang, Y., Sterling, J., Storici, F., Resnick, M. A., & Gordenin, D. A. (2008):** Hypermutability of damaged single-strand DNA formed at double-strand breaks and uncapped telomeres in yeast *Saccharomyces cerevisiae*. *PLoS genetics*, 4(11), e1000264.
85. **Yousaf, A., Jabbar, A., Rajput, N., Memon, A., Shahnawaz, R., Mukhtar, N., ... & Khalil, R. (2019):** Effect of environmental heat stress on performance and carcass yield of broiler chicks. *World's Veterinary Journal*, (1), 26-30.
86. **Yunianto, V. D., Hayashit, K., Kaiwda, S., Ohtsuka, A., & Tomita, Y. (1997):** Effect of environmental temperature on muscle protein turnover and heat production in tube-fed broiler chickens. *British Journal of Nutrition*, 77(6), 897-909.
87. **Zhang, A. W., Lee, B. D., Lee, S. K., Lee, K. W., An, G. H., Song, K. B., & Lee, C. H. (2005):** Effects of yeast (*Saccharomyces cerevisiae*) cell components on growth performance, meat quality, and ileal mucosa development of broiler chicks. *Poultry science*, 84(7), 1015-1021.
88. **Zhou, X., Tian, Z., Wang, Y., & Li, W. (2010):** Effect of treatment with probiotics as water additives on tilapia (*Oreochromis niloticus*) growth performance and immune response. *Fish physiology and biochemistry*, 36, 501-509.